Cortical gating of oropharyngeal sensory stimuli

Karen Wheeler-Hegland1,2*, Teresa Pitts1,2 and Paul W. Davenport2

1 Brain Rehabilitation Research Center, Malcom Randall Veterans Affairs Medical Center, Gainesville, FL, USA
2 Department of Physiological Sciences, University of Florida, Gainesville, FL, USA

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INTRODUCTION

Multiple airway defense reflexes share anatomic substrates contained within the upper airway, including the larynx, pharynx, oral, and nasal cavities. These structures serve as conduits for air and/or food as they participate in the life sustaining functions of respiration and deglutition. The anatomic arrangement of these structures is such that degradation in sensory innervation can impact the generation of precise motor patterns required to swallow or cough safely (Aviv et al., 1997; Addington et al., 1999; Smith Hammond et al., 2001; Teismann et al., 2007) and may lead to uncompensated penetration or aspiration of material to the larynx or lower airways, thereby compromising respiratory function. The timely initiation of a swallow is integral to preventing material from entering the laryngeal vestibule, and the initiation of coughing is integral to ejecting material that may have strayed into the larynx or trachea. Both swallow and cough are dependent upon effective sensory input. Thus, the study of upper airway sensation is an important component in understanding airway defense mechanisms.

Multiple researchers have studied sensory areas that can elicit or modify swallowing and cough using different sensory modalities, including mechanical, gustatory, and thermal stimuli (Doty, 1951; Storey, 1968; Mori and Sakai, 1972; Boushey et al., 1974; Hwang et al., 1984; Sant’Ambrogio et al., 1986; Bradley, 2000; Sciortino et al., 2003; Theurer et al., 2005; Kitagawa et al., 2009). Specifically, the faucial arches and posterior oropharyngeal wall are implicated in swallow initiation, and the laryngeal mucosa for eliciting cough. We have recently shown that air puff stimuli to the posterior oropharynx can induce the urge-to-cough, and in some cases cough (Wheeler-Hegland et al., in press). The pharyngeal branch of the glossopharyngeal and vagus nerves supplies sensory innervation to the posterior oropharyngeal wall (Mu and Sanders, 2000). This pharyngeal plexus of nerve fibers (along with the superior laryngeal nerve), when stimulated, can elicit the swallow reflex (Doty, 1951; Kitagawa et al., 2002) or a cough (Storey, 1968; Sant’Ambrogio et al., 1986). In their detailed study of a human oro- and laryngopharynx, Mu and Sanders (2000) identified areas densely populated with sensory nerve endings including the posterior oropharyngeal wall. They hypothesized that the increased number of nerve fibers in the oropharynx compared with other areas was related to the significance of this area for swallow initiation (Mu and Sanders, 2000).

The study of upper airway sensation in awake humans has traditionally been reported in terms of either motor output or verbal confirmation of sensory perception, making it difficult to differentiate motor, speech and language, or cognitive abilities from sensory parameters. Event-related electroencephalography is a method that does not rely on motor or verbal feedback when studying sensation in humans. This method has high temporal resolution and allows for the scalp recording of neuronal activity that is time-locked to a sensory stimulus, yielding the sensory evoked potential. It has been shown that electrical and mechanical stimuli applied to the lips, buccal cavities, tongue, faucial arches, soft palate, nasal cavity, and nasopharynx will evoke a cortical sensory response (Fujii et al., 1994; Maloney et al., 2000; Hummel et al., 2002; Gow et al., 2004; Nakahara et al., 2004; Yoshida et al., 2006). We have recently shown that air puffs applied to the posterior oropharyngeal wall will elicit
a sensory evoked potential, termed the PSEP (pharyngeal sensory evoked potential), that is characterized by four component peaks: P1, N1, P2, N2, named for their polarity (positive or negative) and relative order (Wheeler-Hegland et al., in press). These peaks are likely analogous to somatosensory evoked potential (SEP) and respiratory-related evoked potential (RREP) peaks with similar latencies and scalp recording locations. The P1 peak is related to the initial arrival of a sensory stimulus at the somatosensory cortex, the N1 and P2 peaks are associated with attention to and processing of the stimulus and the N2 peak is thought to be related to sensory inhibition and decision processing for behavioral responses (Logie et al., 1998; Webster and Colrain, 2000; Crowley and Colrain, 2004; Chan and Davenport, 2008; Ljiffijt et al., 2009).

The term “gating” refers to the ability of the cerebral cortex to include or exclude sensory information from throughput to cortical areas based on its relative redundancy or relevance. Sensory gating out of information is defined as reduced neural activity occurring with increased stimulus redundancy, measured via the scalp sensory evoked potential (Arnfred et al., 2001). Widely studied with the auditory evoked potential (AEP) using a paired-click paradigm (Adler, 1982; Adler et al., 1998), gating has been studied in other sensory systems, including somatosensation and respiration (Arnfred et al., 2001; Chan and Davenport, 2008). In each of these systems, the pairing of two identical stimuli (S1 and S2) results in reduced amplitude (gating out) of S2 such that the ratio of S2/S1 is between 0.33 and 0.59. The SEP P50/RREP P1 peaks and the SEP N100/RREP N1 peaks (analogous to the PSEP P1 and N1, respectively), as well as the AEP peaks P1, N1, and P2, are all considered “gating” peaks because the S2 elicits much smaller amplitude peaks when the paired stimuli are presented within 500 ms of each other.

While the pattern generators that integrate sensory information for airway defense behaviors are in the brainstem (Bolser and Davenport, 2002), the cerebral cortex also plays a role in the genesis of these behaviors (Davenport et al., 2002; Gow et al., 2004; Eccles, 2009). As such, the cortical response to afferent information from the pharynx is important. Oropharyngeal sensory input is specifically important for initiating and modifying the motor events comprising a swallow, and there is some evidence to suggest stimulation of this area can induce coughing. Studying sensation and sensory gating is important to our understanding of these airway defense behaviors. Single swallows, often lasting less than 1000 ms, are readily modified with changes to bolus parameters, such as consistency or volume (Ishida et al., 2002; Lawless et al., 2003). This suggests a need for consistent, online sensory monitoring throughout the duration of a swallow. Therefore, if oropharyngeal stimuli were gated out, the likelihood of losing important information related to the bolus viscosity, size, or location may increase, and hence so might the likelihood of a penetration or aspiration event. As such our overall hypothesis is that central neural gating out of sensory information is absent for stimuli applied to the posterior oropharyngeal wall. Stated another way, oropharyngeal sensory stimuli are gated in at the level of the cerebral cortex. In order to test this hypothesis we identified the following specific aims: (1) to record event-related evoked potentials from the posterior oropharyngeal wall (PSEPs) using paired-pulse air puff stimuli delivered 500 ms apart. It was hypothesized that distinct PSEP waveforms including component peaks P1, N1, P2, and N2 would exist, for the first (S1) and the second (S2) stimulus, and (2) to compare the amplitude of component peaks P1, N1, P2, and N2, from the S1 and S2 PSEP waveforms. It was hypothesized that no significant differences would exist for the amplitude of the S1 and S2 peaks, and that gating ratios (S2/S1) would be greater than 0.60 for the PSEP component peaks.

**MATERIALS AND METHODS**

Twenty-three (7 men, 16 women) healthy volunteers participated in the study. The mean age was 20.65 ± 3.43 years. All subjects self-reported no history of cancer in the head or neck regions, neurologic disease, chronic respiratory disease, or dysphagia. Participants were asked to refrain from caffeine for 12 h prior to participating in the experiment and were not currently smokers. The experimental procedures were explained to the participants upon arrival to the laboratory, and all participants provided written informed consent to participate. The University of Florida Health Science Center Institutional Review Board approved the study.

**PULMONARY FUNCTION TESTING**

All participants were screened with a pulmonary function test to ensure no restrictions or obstructions existed that would indicate airway pathology that would potentially impact sensation. Forced vital capacity (FVC) was measured at least three times for each participant. The participant was instructed to respire normally through a filtered mouthpiece with nose clips in place. Following three to four rest breaths, they were asked to provide a deep inspiration followed by a forced expiration. Instructions were based on the American Thoracic Society standard for spirometry testing (Miller et al., 2005). The forced expired volume within 1 s (FEV1) and the FVC were recorded (Jaeger Toennies, Medizintechnikmit System) and the ratio of FEV1/FVC was used for analysis. All subjects had an FEV1/FVC ratio greater than 75% predicted. The respiratory resistance at 5 Hz (R5) was measured with impulse oscillometry (Jaeger Toennies, Medizintechnikmit System). The mean R5 resistance was 3.00 ± 0.67 cmH2O·L·s, within the predicted normal range for all participants.

**SUBJECT PREPARATION AND PROTOCOL**

A 32-electrode Neuroscan Quickcap™ based on the International 10–20 system was positioned on the participant’s head and connected to the SynAmps2 Neuroscan System. Conducting gel was applied through each electrode in order to establish scalp contact and maintain impedance levels below 5 kΩ. Bipolar electrodes were placed on the skin above and below the left eye for recording vertical electro-oculogram (VEOG) activity. Synamps2 amplifiers (Neuroscan, El Paso, TX, USA) and SCAN version 4.3 acquisition software (Neuroscan, El Paso, TX, USA) were used to record the EEG signal onto a desktop computer. The sampling rate was set to 1000 Hz per channel with a recording bandpass of DC to 200 Hz. The EEG activity was referenced to linked earlobes. SCAN version 4.3 analysis software (Neuroscan, El Paso, TX, USA) was used for data analysis.

Participants were instructed to relax and sit comfortably in a chair with the neck, back, and arms supported. A mouthpiece with a polyethylene tube was placed in the mouth, and a flexible laryngoscope...
was inserted through the tube (**Figure 1**). The laryngoscopic images were displayed, but not recorded, on a computer screen. Both the laryngoscope and computer were components of the JEDMED StroboCAM II® system (JEDMED Instrument Co., St Louis, MO, USA). In this manner, the laryngoscope allowed for visualization and verification of tube placement for air puff delivery. The laryngoscope itself was covered with a hygienic sheath (Slide-On® Sheath for Sensory Testing, Medtronic Xomed, Inc., Jacksonville, FL, USA) that has a small port through which the air puffs were delivered. The port was connected to an air tank connected to a solenoid valve that delivered air puffs to the laryngoscope tube. The air tank was in-series with a digital manometer (Fluke 713 30G, John Fluke MFG Co-Inc, Everett, WA, USA), allowing for control of air puff pressure. A manual trigger system that provided an electrical output was used to initiate the data sample collected by the computer simultaneously with air puff delivery.

Air puffs were delivered in pairs with an inter-stimulus interval (ISI) of 500 ms. This interval was chosen based on the work of Arnfred et al. (2001) that identified 500 ms as being the optimal paired stimulus interval for measuring sensory gating. Four sets of 50 air puff pairs were delivered, yielding a total of 200 stimuli for stimulus 1 (S1) and stimulus 2 (S2). Between each air puff pair (within sets of 50) there was a minimum of 2-s interval. This took approximately 3–4 min per set; participants rested for at least 2 min between sets and were allowed to drink water at that time, if desired.

**FIGURE 1** Mouthpiece with polyurethane tubing used for placement of the flexible laryngoscope with the air puff delivery port. The laryngoscope was inserted posteriorly toward the oropharyngeal wall, therefore oriented at an approximately 90° angle.

**EXPERIMENTAL PROTOCOL**

Air puff pairs were delivered through the laryngoscope port, inserted and anchored through the mouthpiece. In order to place the delivery port, participants were instructed to relax their jaw and tongue, and to breathe through the mouth. Participants were then asked to sustain “ah” which raised the soft palate and allowed for insertion of the tube through the posterior oral cavity and into the oropharyngeal region. The delivery port was situated immediately in front of, but not touching, the paramedial posterior oropharyngeal wall, within approximately 5 mm of midline on either side. Placement was verified visually on the computer screen. Once in place, preliminary air puffs were delivered in order to establish that the participant could feel the air puff specifically on the posterior oropharyngeal wall. It was not acceptable to feel generalized pressure in the throat, and the air puff pressure was adjusted until subjects indicated they felt a discrete air puff stimulus on the posterior oropharyngeal wall. The duration of each air puff pulse was 150 ms; pressure of the air puffs was recorded for each subject, and ranged from 15 to 30 cmH2O at the air tank. As pressure dissipates as it travels through the tube, the pressure at the delivery port ranged between 1 and 4 cmH2O.

**DATA ANALYSIS**

Five-hundred millisecond epochs (100 ms pre-stimulus and 400 ms post stimulus) were sampled when the air puff stimulus was triggered for both S1 and S2. During offline analysis using SCAN 4.3 software (Neuroscan, El Paso, TX, USA) each S1 and S2 data frame was reviewed and the inclusion criteria for epochs were: (1) no VEOG eyeblink activity, and (2) no change of EEG activity exceeding 50 μV. Responses to stimuli that were confounded by artifacts were excluded from analysis. A minimum of 190 air puff epochs were averaged for both stimuli to obtain the PSEP waveforms. The peak latencies were measured from the time of S1 and S2 onset, and amplitudes were measured from baseline to peak for each component peak.

Sensory gating is based on peak amplitude and accuracy is dependent upon measurement of the same peak for the S1 and S2 waveform. As such, a latency criterion of ±10 ms between the S1 and S2 peaks was established. Identification of the component peaks has been described in detail elsewhere (Wheeler-Hegland et al., in press). Briefly, the averaged S1 and S2 waveforms for each subject were displayed concurrently on a computer screen. As well, along with the waveforms a 2-D head model corresponding temporally to the waveform and depicting all recording electrodes was displayed (**Figure 2**) using source localization software (Scan 4.3). The software was used to determine EEG amplitude at and between the electrodes. Concurrent visualization of the S1 and S2 waveforms and 2-D head models allowed for identification of the positive and negative component peaks. Once a peak was identified for each stimulus, the recording electrode closest to the area of greatest positive or negative EEG amplitude was identified. This is termed the “hot spot” electrode. If the hot spot electrode was different between S1 and S2 the closest electrode to the hot spot that was common between the two stimuli was chosen.

Descriptive statistics including mean values and standard deviations were used to describe the latency, amplitude, and S2/S1 ratio data. A repeated measures analysis of variance was used to test for significant differences in the amplitude of S2 and S1 responses for each of the four component peaks.
RESULTS

Each of the component peaks P1, N1, P2, and N2 were present for both the S1 and S2 stimuli in all subjects. Their mean latencies and amplitudes are in Table 1. Results of repeated measures ANOVA with within subject factors stimulus (S1 or S2) and peak (P1, N1, P2, N2) showed non-significant differences between the S1 and S2 peak amplitudes ($F = 1.428$, df = 22.3, $p = 0.243$).

Gating ratios were computed by dividing the amplitude of S2 by that of S1 (S2/S1). A ratio of less than 0.60 was established as criteria for S2 to be considered gated (Chan and Davenport, 2008). Mean gating ratio and standard error for P1 is 0.92 (0.13), for N1 is 1.12 (0.11), for P2 is 1.09 (0.09), and for N2 is 0.96 (0.10). Figure 3 depicts the gating ratio of each peak for individual subjects. In total, the P1 peak was considered gated in 7 of 23 subjects, the N1 peak was gated in 5 of 23 subjects, and P2 and N2 were each gated in 3 of 23 subjects. When gating did occur, there was no pattern observed between the peaks. For example, gating of P1 did not necessarily mean that N1 or other subsequent peaks were also gated.

DISCUSSION

Pharyngeal sensory evoked potential P1, N1, P2, and N2 peaks were present in response to each stimulus in a paired-air puff paradigm with an ISI of 500 ms. The S2 response failed to yield significantly different amplitude versus the S1 response amplitude in the P1, N1, P2, or N2 peaks. The air puff parameters for S1 and S2 were constant indicating that these results were not a function of variations in stimulus pressure intensity or duration. Comparison and analysis of the amplitudes of the S1 and S2 peaks shows that repetitive redundant input to the oropharynx is not gated out. These results are inconsistent with those
previously reported for other sensory modalities, thus supporting our *a priori* hypothesis that sensory central neural gating out would be absent with identical, paired-pulse air puff stimuli delivered to the oropharynx. These differences are not likely due to protocol deviations as the spacing of stimuli was as recommended by Arnfred et al. (2001) examining the effect on ISI on gating relationships. Their study found 500 ms ISI to be most effective in terms of the degree of gating present in both somatosensory and auditory modalities. Chan and Davenport (2008) also used 500 ms ISI and showed gating is present for the RREP and mouth evoked potential recorded with air puffs delivered to the inner surface of the cheek (buccal cavity).

Gating deficits, a term used to indicate a lack of reduction in evoked potential peak amplitude for S2 versus S1 redundant stimuli, exist in various pathologies including some strokes (Staines et al., 2002), schizophrenia (Adler, 1982), and obsessive–compulsive disorder (Nanbu et al., 2010). Gating deficits are hypothesized to indicate abnormal influx of redundant sensory information that impedes the brain’s ability to efficiently process sensory input, and may impede higher cognitive processes (Lijffijt et al., 2009). However, it has been shown that healthy people can endogenously modulate gating under certain conditions. For example, directing research participants to attend to specifically to the S2 stimulus can result in disinhibition of the S2 response (Guterman et al., 1992; Jerger et al., 1992; White and Yee, 1997; Webster and Colrain, 2000). This effect is dependent upon the type of attentional resources required, as simply directing a participant to attend to the presence of stimuli does not affect gating ratios (Harver et al., 1995; Chan and Davenport, 2009). Therefore, it is necessary to require some higher level of processing in order to effectively disinhibit the S2 response.

**Table 1 | Mean values and standard errors (SE) for latency and amplitude of the S1 and S2 stimuli.**

<table>
<thead>
<tr>
<th></th>
<th>Latency (ms) Mean (SE)</th>
<th>Amplitude (μV) Mean (SE)</th>
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</thead>
<tbody>
<tr>
<td></td>
<td>S1</td>
<td>S2</td>
</tr>
<tr>
<td>P1</td>
<td>57.96 (2.26)</td>
<td>59.22 (2.38)</td>
</tr>
<tr>
<td>N1</td>
<td>90.00 (3.91)</td>
<td>92.74 (3.86)</td>
</tr>
<tr>
<td>P2</td>
<td>122.00 (4.65)</td>
<td>124.26 (4.92)</td>
</tr>
<tr>
<td>N2</td>
<td>165.13 (6.12)</td>
<td>166.13 (6.10)</td>
</tr>
</tbody>
</table>

Participants in the current study were not given any instructions on whether to attend to the stimuli, and all watched a movie of their choice during the experiment. However, we have reported for a subpopulation of the participants in the current study there was a significant urge-to-cough during the experiment (Wheeler-Hegland et al., in press). While it was not measured explicitly in the current study, previous research tells us that repetitive air puffs delivered to the posterior faucial arches can induce the urge-to-swallow or swallowing (Theurer et al., 2005). Thus, there was likely some degree of generalized attention to the sensations generated via air puff stimuli. Guterman et al. (1992) propose that attention as it relates to gating can serve to facilitate cortical responsiveness to incoming stimuli. Chan and Davenport (2009) demonstrated for both the mouth and respiratory evoked potential that ratios in an “attend” to the stimulus condition were still gated. Hence it is possible that for any stimulus delivered to the oropharyngeal region, whether redundant or novel, there may be a level of attention related to responsiveness for airway defense (e.g., swallowing or cough), it is unlikely that attention to the stimuli accounts for the overall lack of gating found in this study.

Attention should also be given to the degree of variability in amplitude measures and the possibility of type 2 error, or failing to reject the null hypothesis (that no differences would be detected) when significant differences do exist. Typically for studies on sensory gating the primary outcome measure is the S2/S1 (gating) ratio with criterion set at 0.6 or less for a stimulus to be considered gated. The repeated measures ANOVA test was included in this analysis in the case ratios were very close to that ratio criteria there would be some measure of determining whether a significant difference existed between the two stimuli. Therefore, while the possibility of type II error cannot be ruled-out, based on degree to which the gating ratios are greater than 0.6 there is confidence in the conclusion that these are peaks are not gated. To further check this, a one-sample *t*-test was completed for gating ratios (mean = 1.02, SD = 0.52) against S2/S1 = 0.60 that yielded a significant difference (*t* = 7.742, df = 91, *p* < 0.000), indicating the mean gating ratio was greater than 0.6.

Our method of identifying S1 and S2 component peaks is novel in that we did not identify a recording electrode *a priori* and instead identified the hot spot electrode for each participant as part of data analysis. We developed this method of peak identification because of the availability of 2-D mapping software that allowed for visualization of dipole evolution, from initial creation to the

![FIGURE 3 | Gating ratios of each peak for all individual subjects.](image-url) The figure legend is on the right-hand side. Each shape represents one of the four peaks.
peak. It was therefore possible to identify the strongest recording location within a latency window for each dipole, accounting for individual differences in scalp topography in our analyses. Studies including the Arnfred study, and the Chan and Davenport studies chose a priori recording electrodes and then identified the peaks at those electrodes within a specific latency window. What is ultimately important is that the same recording electrodes were chosen from both the first and second stimulus within the same latency windows, ensuring comparison of the same peak from S1 and S2, which is fundamental to the measurement of sensory gating.

Sensory gating is thought to be the result of an effect for the S1 stimulus on subcortical structures, notably the thalamus (Kaji, 2001), and possibly its connections with the hippocampus and prefrontal cortex (Tregellas et al., 2007), ultimately acting on the throughput of the S2 stimulus. Event-related EEG recording from the scalp cannot determine the precise neuroanatomical substrates due to poor spatial resolution, thus rendering it difficult to make any conclusion regarding the structure(s) that regulate(s) gating of oropharyngeal sensory information. Presently it is only possible to hypothesize based on results from imaging studies. Multiple imaging studies have shown diffuse areas of cortical activation during swallowing (Hamdy et al., 1999; Mosier et al., 1999; Martin et al., 2001; Toogood et al., 2005), swallow-related behaviors (Martin et al., 2004; Lowell et al., 2008; Malandraki et al., 2009), coughing (Simonyan et al., 2007), urge-to-cough (Mazzone et al., 2007), and oral air puff stimulation (Lowell et al., 2008; Sörös et al., 2008). These findings provide several possible neuroanatomical substrates for the findings of the current study. Operating under the hypothesis that the thalamus is a key structure for sensory gating, it is interesting that several thalamic nuclei are active during swallowing coughing, and oral air puff stimulation. Consider as well the functional cortical model of swallowing substrates proposed by Mosier and Bereznyay (2001), consisting of five modules and two parallel loops. Within their model is the IGF-S2-CC-BGTHAL module consisting of the inferior frontal gyrus, secondary sensory cortex, corpus callosum, and basal ganglia and thalamus. This group of structures is thought to integrate sensory information about the bolus with some internal representation of swallowing movement. It would therefore fit the functionality of this module and with the thalamus gating hypothesis if gating in or out occurred as part of the decision-making process. Future studies incorporating EEG and functional magnetic imaging, or magnetoencephalography would enable anatomical correlation and electrophysiological data to be collected simultaneously in order to glean more specific information regarding the anatomy and physiology of oropharyngeal sensory gating.

For some patients with dysphagia (disordered swallowing) or dystussia (disordered coughing) gating out of oropharyngeal sensory information may be related to dysfunctional formation or modulation of the motor output plan. For example, if during the course of a meal, pharyngeal sensory information is gated out at a level that does not allow for normal initiation of swallow motor output, perhaps a progressive delay in swallow initiation would occur and result in airway compromise. This is highly speculative, and future research should examine oropharyngeal sensory gating in disordered populations in order to ascertain: (1) pharyngeal gating characteristics, and (2) potential relationship to swallow dysfunction and airway compromise.

REFERENCES


Conflict of Interest Statement: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.